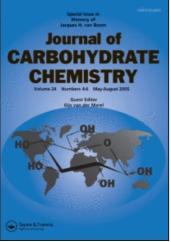
This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

Synthetic Studies on Sialoglycoconjugates 36: α -Selective Glycoside Synthesis of *N*-Acetylneuraminic Acid with the Secondary Hydroxyl Group in D-Glucofyranose, 2-Acetamido-2-deoxy-D-glucopyranose and D-Galactopyranose Derivatives

Akira Hasegawa^a; Masayuki Ogawa^a; Yukiya Kojima^a; Makoto Kiso^a ^a Department of Applied Bioorganic Chemistry, Gifu University, Gifu, Japan

To cite this Article Hasegawa, Akira , Ogawa, Masayuki , Kojima, Yukiya and Kiso, Makoto(1992) 'Synthetic Studies on Sialoglycoconjugates 36: α -Selective Glycoside Synthesis of *N*-Acetylneuraminic Acid with the Secondary Hydroxyl Group in D-Glucofyranose, 2-Acetamido-2-deoxy-D-glucopyranose and D-Galactopyranose Derivatives', Journal of Carbohydrate Chemistry, 11: 3, 333 – 341

To link to this Article: DOI: 10.1080/07328309208017997 URL: http://dx.doi.org/10.1080/07328309208017997

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

SYNTHETIC STUDIES ON SIALOGLYCOCONJUGATES 36: α-SELECTIVE GLYCOSIDE SYNTHESIS OF *N*-ACETYLNEURAMINIC ACID WITH THE SECONDARY HYDROXYL GROUP IN D-GLUCOPYRANOSE, 2-ACETAMIDO-2-DEOXY-D-GLUCOPYRANOSE AND D-GALACTOPYRANOSE DERIVATIVES

Akira Hasegawa, Masayuki Ogawa, Yukiya Kojima, and Makoto Kiso

Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11, Japan

Received November 4, 1991 - Final form December 30, 1991

ABSTRACT

Coupling of the secondary hydroxyl group in the 6-O-benzoyl derivatives 3, 5 and 8, prepared from 2-(trimethylsilyl)ethyl β -D-glucopyranoside (2), 2-acetamido-2-deoxy- β -D-glucopyranoside (4), and 2-acetamido-2-deoxy- β -D-galactopyranoside (6), with methyl (methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-2-thio-D-glycero- α -D-galacto-2-nonulopyranosid)onate (1) as the glycosyl donor in acetonitrile in the presence of dimethyl(methylthio)sulfonium triflate (DMTST) as a glycosyl promoter, exclusively gave the α -glycosides at C-2 in 3, at C-3 in 5 and 8, respectively.

INTRODUCTION

Sialic acids are well known as constituents of glycoproteins and glycolipids of cell membranes, and they play important roles in various biological processes. As far as we know, sialic acids are linked in α -configuration at C-3, -4 and -6 of the Gal, at C-6 of the Glc, at C-6 of the GlcNAc and GalNAc, and at C-8 of the sialic acid residue in sialoglycoconjugates. Therefore, a facile α -selective glycoside synthesis of sialic acids, especially with the secondary hydroxyl groups of sugar derivative, is critically important in order to investigate the functions of sialoglycoconjugates at the molecular level. Recently, new efforts^{1,2} have been made at obtaining mainly α -glycosides of sialic acid using 3-substituted Neu5Ac derivatives as glycosyl donors. We have developed³ a facile α -stereoselective glycosylation of sialic acids by using 2-thioglycosides of sialic acids as the glycosyl donor and suitably protected acceptors, with dimethyl(methylthio)sulfonium triflate⁴ (DMTST) or N-iodosuccinimide (NIS)-trifluoromethanesulfonic acid⁵ (Tf-OH) as the glycosyl promoter in acetonitrile under kinetically controlled conditions. The α -glycosides of sialic acids thus obtained were effectively employed⁶ as the building blocks for the following ganglioside synthesis. As a part of our continuing efforts on the synthesis of α -glycosides of sialic acid and sialoglycolipids, we describe here the synthesis of sialosyl α -(2 \rightarrow 2)-D-glucopyranose and sialosyl α -(2 \rightarrow 3)-2-acetamido-2-deoxy -D-gluco- and D-galactopyranoses derivatives.

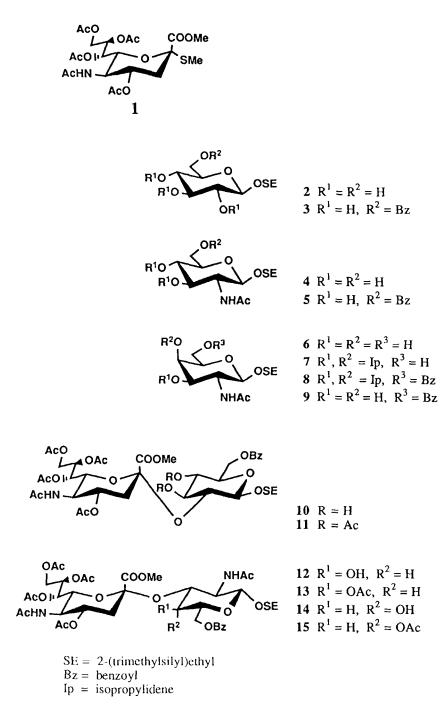
RESULTS AND DISCUSSION

For the synthesis of the desired α -glycosides of Neu5Ac, we have chosen the 6-Obenzoylated derivatives 3, 5, and 9 of 2-(trimethylsilyl)ethyl β -D-glucopyranoside⁷ (2), 2-acetamido-2-deoxy- β -D-glucopyranoside⁸ (4), and 2-acetamido-2-deoxy- β -D-galactopyranoside⁹ (6) as the glycosyl acceptor. We have employed methyl (methyl 5acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-2-thio-D-glycero- α -D-galacto-2-nonulopyranosid)onate¹⁰ (1) as the glycosyl donor.

Treatment of 2 or 4 with benzoyl chloride in pyridine-dichloromethane at -50 °C gave the corresponding 6-O-benzoyl derivatives 3 and 5 in 60% yields, respectively.

Treatment of 6 with 2,2-dimethoxypropane in N,N-dimethylformamide in the presence of p-toluenesulfonic acid monohydrate for 2 h at 80 °C, gave the 3,4-O-isopropylidene derivative 7 in 80% yield, which, on O-benzoylation and subsequent hydrolysis of the isopropylidene group with 80% aqueous acetic acid for 2 h at 45 °C, afforded 2-(trimethylsilyl)ethyl 2-acetamido-6-O-benzoyl-2-deoxy- β -D-galactopyranoside (9) in high yield.

The glycosylation of 3 with the methyl α -2-thioglycoside 1 (2.0 equiv. relative to the acceptor) of Neu5Ac in acetonitrile for 15 h at -15-20 °C in the presence of DMTST, gave the expected α -glycoside 10 of Neu5Ac, 2-(trimethylsilyl)ethyl O-(methyl 5acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -galacto-2-nonulo-pyranosylonate)-(2 \rightarrow 2)-6-O-benzoyl- β -D-glucopyranoside (10) in 48% yield. The observed chemical shifts and coupling constants (δ 2.91, J3_a,3e = 12.5 Hz, J3_e,4 = 4.4 Hz, δ 4.98, and δ 5.40, J7,8 = 7.8 Hz) for H-3e, H-4, and H-7 in Neu5Ac moiety are characteristic for α -glycosidic linkages.²,4b,11,12 The linkage position of the Neu5Ac



unit to the Glc residue was unambiguously proved from the ¹H NMR data of the acetvlated derivative 11. Significant signals of 11 were a one-proton doublet of doublets at δ 4.28 (J_{1,2} = 8.1 Hz, J_{2,3} = 8.4 Hz, H-2) and a one-proton doublet at δ 4.41 (H-1) in the Glc residue, indicating the linkage position assigned. Other ¹H NMR data are given in the Experimental Section and are consistent with the structure. In the same way, when treated with the glycosyl acceptors 5 and 9, compound 1 yielded the corresponding α glycosides 12 and 14 at O-3 in the acceptor moiety in 49 and 46% yields, respectively; no β -glycoside and no position isomer of Neu5Ac was isolated. Acetylation of the glycosides 12 and 14 thus obtained gave the acetates 13 and 15. The structures of the glycosides 13 and 15 were determined from the ^{1}H NMR data; the observed chemical shifts and coupling constants of the sialic acid units for H-3e (δ 2.58, J_{3a,3e} = 12.8 Hz, $J_{3e,4} = 4.4$ Hz for 13; δ 2.66, $J_{3a,3e} = 12.5$ Hz, $J_{3e,4} = 4.4$ Hz, for 15) and for H-4 $(\delta 4.91, J_{3a,4} = 12.5 \text{ Hz for } 13; \delta 4.80, J_{3a,4} = 12.4 \text{ Hz for } 15)$, are of α -anomeric configuration; H-4 for 13 appeared at δ 5.09 (t, J3,4 = J4,5 = 8.1 Hz) and for 15 at δ 4.95 (broad d), indicating the linkage position of the Neu5Ac residue in 13 and 15 to be O-3. Other ¹H NMR data are consistent with structures 13 and 15, respectively.

In conclusion, α -stereoselective glycosylation of Neu5Ac has been also observed by using the methyl 2-thioglycoside 1 of Neu5Ac as the glycosyl donor and the 6-Obenzoyl derivatives 3, 5, and 9 of 2-(trimethylsilyl)ethyl β -D-glucopyranoside and 2acetamido-2-deoxy- β -D-hexopyranosides) with DMTST in acetonitrile under kinetically controlled conditions. The α -glycosides described herein could be used as intermediates suitable for the neoganglioside synthesis, and they are also inportant as building units for sialoglycoconjugate synthesis.

EXPERIMENTAL

General Procedures. Specific rotations were determined with a Union PM-201 polarimeter at 25 °C, and IR spectra were recorded with a Jasco A-100 spectrophotometer. ¹H NMR spectra were recorded with a JEOL JNM-GX 270 spectrometer. Preparative chromatography was performed on silica gel (Wako Chemical Co., 200 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

2-(Trimethylsilyl)ethyl 6-O-Benzoyl- β -D-glucopyranoside (3). To a solution of 2-(trimethylsilyl)ethyl β -D-glucopyranoside⁷ (2; 1.01 g, 3.6 mmol) in pyridine (10 mL) and dichloromethane (10 mL), cooled to -50 °C, was added benzoyl chloride (0.63 mL, 5.4 mmol), and the mixture was stirred for 15 min at -30 °C.

The reaction was monitored by TLC and, when complete, methanol (1 mL) was added, and the mixture was concentrated. Column chromatography (10:1 ethyl acetate-

hexane) of the residue on silica gel (60 g) gave **3** (830 mg, 60%) as an amorphous mass: $[\alpha]_D$ -50.5° (*c* 0.95, CHCl₃); ¹H NMR (CDCl₃) δ 1.01 (m, 2H, Me₃SiCH₂CH₂), 4.36 (d, 1H, J_{1,2} = 7.7 Hz, H-1), 4.49 (dd, 1H, J_{5,6}' = 6.2 Hz, J_{6,6}' = 12.1 Hz, H-6'), 4.72 (dd, 1H, J_{5,6} = 1.7 Hz, H-6), and 7.42-8.08 (m, 5H, Ph).

Anal. Calcd for C18H28O7Si (384.5): C, 56.22; H, 7.34. Found: C, 56.31; H, 7.50.

2-(Trimethylsilyl)ethyl 2-Acetamido-6-O-benzoyl-2-deoxy- β -D-glucopyranoside (5). To a solution of 4⁸ (2.0 g, 6.2 mmol) in pyridine (10 mL) and dichloromethane (10 mL), cooled to -50 °C, was added benzoyl chloride (1.1 mL, 9.35 mmol), and the mixture was stirred for 20 min at -50 °C. MeOH (1 mL) was added, and the mixture was concentrated. Column chromatography (2:1 ethyl acetate-hexane) of the residue on silica gel (70 g) gave 5 (1.56 g, 59%) as an amorphous mass: [α]_D-49.0° (*c* 0.8, CHCl₃); IR (KBr) 3600-3300 (OH, NH), 1730 and 1280 (ester), 1660 and 1560 (amide), 860 and 840 (TMS), and 710 cm⁻¹ (Ph); ¹H NMR (1:1 CD₃OD-CDCl₃) δ 0.98 (m, 2H, Me₃SiCH₂CH₂), 2.08 (s, 3H, AcN), 3.96 (m, 1H, Me₃SiCH₂CH₂), 4.48-4.61 (m, 2H, H-6,6'), 4.79 (d, 1H, J_{1,2} = 11.0 Hz, H-1), and 7.40-8.07 (m, 5H, Ph).

Anal. Calcd for C₂₀H₃₁NO₇Si (425.6): C, 56.42; H, 7.34; N, 3.29. Found: C, 56.43; H, 7.41; N, 3.18.

2-(Trimethylsilyl)ethyl 2-Acetamido-2-deoxy-3,4-O-isopropylidene- β -D-galactopyranoside (7). To a solution of 6⁹ (1.0 g, 3.1 mmol) in *N*,*N*dimethylformamide (DMF; 10 mL) were added 2,2-dimethoxypropare (0.77 mL, 6 mmol) and *p*-toluenesulfonic acid monohydrate (10 mg), and the mixture was stirred for 2 h at 80 °C and neutralized with Amberlite IR-410 (OH⁻) resin. The resin was filtered off and washed with methanol, and the combined filtrate and washings were concentrated. Column chromatography (2:1 ethyl acetate-hexane) of the residue on silica gel (100 g) gave 7 (980 mg, 81%) as an amorphous mass: $[\alpha]_D + 1.6^\circ$ (*c* 2.0, CHCl3); IR (KBr) 3600-3300 (OH, NH), 1660 and 1550 (amide), and 840 cm⁻¹ (TMS, Me₂C); ¹H NMR (CDCl₃) δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.32, 1.51 (2s, 6H, Me₂C), 1.98 (s, 3H, AcN), 3.54 (m, 1H, Me₃SiCH₂CH₂), 4.75 (dd, 1H, J₃,4 = 5.5 Hz, J₄,5 = 2.6 Hz, H-4), 5.00 (d, 1H, J₁,2 = 8.4 Hz, H-1), and 5.94 (d, 1H, NH).

Anal. Calcd for C₁₆H₃₂NO₆Si (347.5): C, 55.30; H, 9.28; N, 4.03. Found: C, 55.28; H, 9.40; N, 4.01.

2-(Trimethylsilyl)ethyl 2-Acetamido-6-O-benzoyl-2-deoxy-3,4-O-isopropylidene- β -D-galactopyranoside (8). To a solution of 7 (710 mg, 2.04 mmol) in pyridine (10 mL), cooled to -20 °C, was added benzoyl chloride (0.4 mL, 3.4 mmol), and the mixture was stirred for 15 min at -5 °C and then worked-up, as described for 3, to give 8 (770 mg, 84%) as an amorphous mass: $[\alpha]_D$ +22.5° (c 1.1, CHCl3); ¹H NMR (CDCl₃) δ 0.96 (m, 2H, Me₃SiCH₂CH₂), 1.37, 1.57 (2s, 6H, Me₂C), 2.01 (s, 3H, AcN), 3.59, 3.98 (2m, 2H, Me₃SiCH₂CH₂), 5.03 (d, 1H, J_{1,2} = 8.4 Hz, H-1), 6.64 (d, 1H, J_{2,NH} = 7.3 Hz, NH), and 7.43-8.09 (m, 5H, Ph).

Anal. Calcd for C23H35NO7Si (451.6): C, 61.17; H, 7.81; N, 3.10. Found: C, 61.22; H, 7.98; N, 3.15.

2-(Trimethylsilyl)ethyl 2-Acetamido-6-O-benzoyl-2-deoxy- β -Dgalactopyranoside (9). A solution of 8 (770 mg, 1.7 mmol) in aqueous 80% acetic acid (15 mL) was heated for 2 h at 45 °C, and concentrated. Column chromatography (50:1 dichloromethane-methanol) of the residue on silica gel (80 g) gave 9 (730 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -35.0° (c 0.5, CHCl3); IR (KBr) 3700-3200 (OH, NH), 1710 and 1250 (ester), 1630 and 1550 (amide), 850 and 830 (TMS), and 710 cm-1 (Ph); ¹H NMR (CDCl3) δ 0.93 (m, 2H, Me3SiCH₂CH₂), 2.02 (s, 3H, AcN), 3.61 (m, 1H, Me3SiCH₂CH₂), 4.50 (d, 1H, J_{1,2} = 8.4 Hz, H-1), and 7.45-8.08 (m, 5H, Ph).

Anal. Calcd for C₂₀H₃₁NO₇Si (425.6): C, 56.42; H, 7.34; N, 3.29. Found: C, 56.35; H, 7.36; N, 3.19.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4,7,8,9-tetra-Oacetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)- $(2\rightarrow 2)$ -6-O-benzoyl- β -D-glucopyranoside (10). To a solution of 3 (1.5 g, 3.9 mmol) and 1¹⁰ (4.1 g, 7.9 mmol) in acetonitrile (15 mL) were added powdered molecular sieves 3Å (5.0 g), and the mixture was stirred for 6 h at room temperature, then cooled to -20 °C. A mixture of dimethyl(methylthio)sulfonium triflate (DMTST; 4 g) and molecular sieves 3Å (4 g) was added, the mixture was stirred for 24 h at -20 °C, and the reaction was monitored by TLC. The precipitate was collected and washed with dichloromethane, and the combined filtrate and washings were washed with M sodium carbonate and water, dried (Na2SO4), and concentrated. Column chromatography (2:1 ethyl acetate-hexane) of the residue on silica gel (250 g) gave 10 (1.6 g; 48%) as an amorphous mass: [\alpha]_ -17.8° (c 1.0, CHCl3); IR (KBr) 3600-3300 (OH, NH), 1750 and 1230 (ester), 1660 and 1550 (amide), 860 and 840 (TMS), and 710 cm⁻¹ (Ph); ¹H NMR (CDCl₃) Neu5Ac unit δ 1.94 (s, 3H, AcN), 2.03, 2.07, 2.17, 2.18 (4s, 12H, 4AcO), 2.91 (dd, 1H, $J_{3a,3e} = 12.5$ Hz, $J_{3e,4} = 4.4$ Hz, H-3e), 3.82 (s, 3H, MeO), 4.02 (q, 1H, $J_{4,5} = J_{5,6} = J_{5,NH} = 10.1$ Hz, H-5), 4.13 (dd, 1H, $J_{8,9'} = 5.5$ Hz, $J_{9,9'} = 12.5 Hz, H-9'$, 4.43 (dd, 1H, $J_{8,9} = 2.6 Hz, H-9$), 4.98 (m, 1H, H-4), 5.40 (dd, 1H, J_{6.7} = 2.0 Hz, J_{7.8} = 7.8 Hz, H-7), 5.44 (m, 1H, H-8), and 5.54 (d, 1H, NH); Glc unit δ 0.87-1.05 (m, 2H, Me₃SiCH₂CH₂), 4.35 (d, 1H, J_{1,2} = 7.7 Hz, H-1), and 7.42-8.13 (m, 5H, Ph).

Anal. Calcd for C38H55NO19Si (857.9): C, 53.20; H, 6.46; N, 1.63. Found: C, 53.31; H, 6.70; N, 1.62.

A sample of 10 (92 mg, 0.11 mmol) was acetylated with acetic anhydride (1 mL)pyridine (2 mL) overnight at room temperature. Column chromatography (80:1 dichloromethane-methanol) of the product on silica gel (30 g) gave 11 (100 mg, quantitative) as an amorphous mass: $[\alpha]_D$ -6.5° (*c* 1.1, CHCl3); ¹H NMR (CDCl3) Neu5Ac unit δ 1.87 (s, 3H, AcN), 2.30 (dd, 1H, J_{3a,3e} = 12.5 Hz, J_{3e,4} = 4.8 Hz, H-3e), 3.83 (s, 3H, MeO), 4.50 (dd, 1H, J_{8,9}' = 2.6 Hz, J_{9,9}' = 12.1 Hz, H-9'), 4.87 (m, 1H, H-4), 5.40 (dd, 1H, J_{6,7} = 2.5 Hz, J_{7,8} = 9.3 Hz, H-7), and 5.52 (m, 1H, H-8); Glc unit δ 0.96 (m, 2H, Me₃SiCH₂CH₂), 3.60 (m, 1H, Me₃SiCH₂CH₂), 4.25 (dd, 1H, J_{1,2} = 8.1 Hz, J_{2,3} = 8.4 Hz, H-2), 4.41 (d, 1H, H-1), 5.20 (m, 2H, H-3,4), and 7.31-8.13 (m, 5H, Ph); *O*-acetyl groups 2.03, 2.05, 2.06, 2.10, 2.18, 2.27 (6s, 18H, 6AcO).

Anal. Calcd for C42H59NO21Si (942.0): C, 53.55; H, 6.31; N, 1.49. Found: C, 53.64; H, 6.39; N, 1.43.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4,7,8,9-tetra-Oacetyl-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonate)- $(2 \rightarrow 3)$ -2-acetamido-6-O-benzoyl-2-deoxy-β-D-glucopyranoside (12). Glycosylation of 5 (460 mg, 1.08 mmol) with 1 (1.2 g, 2.3 mmol) in acetonitrile (6 mL) in the presence of DMTST (1.02 g) and molecular sieves 3Å (2.0 g), as described for the synthesis of 10, gave 12 (465 mg, 49%) as an amorphous mass: $[\alpha]_D$ -26.7° (c 0.9, CHCl3); ¹H NMR (CDCl3) Neu5Ac unit δ 2.68 (dd, 1H, J_{3a,3e} = 12.5 Hz, J_{3e,4} = 4.6 Hz, H-3e), 3.76 (s, 3H, MeO), 4.37 (m, 1H, H-9), 4.81 (m, 1H, H-4), 5.29 (dd, 1H, J_{6,7} = 2.0 Hz, J_{7,8} = 8.5 Hz, H-7), and 5.67 (m, 1H, H-8); GlcNAc unit δ 0.98 (m, 2H, Me3SiCH₂CH₂), 4.52 (d, 1H, J_{1,2} = 8.4 Hz, H-1), and 7.41-8.06 (m, 5H, Ph); other groups δ 1.86, 2.02 (2s, 6H, 2AcN), 2.05, 2.13, 2.16, 2.21 (4s, 12H, 4AcO), and 5.40, 6.59 (2d, 2H, 2NH).

Anal. Calcd for C40H58N2O19Si (899.0): C, 53.44; H, 6.50; N, 3.12. Found: C, 53.39; H, 6.34; N, 3.24.

A sample of 12 (150 mg, 0.17 mmol) was acetylated with acetic anhydride (2 mL)pyridine (4 mL), as described for 11, to give 13 (143 mg, 87%) as an amorphous mass: $[\alpha]_D -11.1^\circ$ (*c* 1.0, CHCl₃); ¹H NMR (CDCl₃) Neu5Ac unit δ 2.58 (dd, 1H, J_{3a,3e} = 12.8 Hz, J_{3e,4} = 4.4 Hz, H-3e), 3.80 (s, 3H, MeO), 4.91 (m, 1H, J_{3a,4} = 12.5 Hz, H-4), 5.31 (dd, 1H, J_{6,7} = 1.2 Hz, J_{7,8} = 7.9 Hz, H-7), and 5.54 (m, 1H, H-8); GlcNAc unit δ 0.97 (m, 2H, Me₃SiCH₂CH₂), 3.61 (m, 1H, Me₃SiCH₂CH₂), 4.48 (d, 1H, J_{1,2} = 7.7 Hz, H-1), 5.09 (t, 1H, J_{3,4} = J_{4,5} = 8.1 Hz, H-4), and 7.34-8.04 (m, 5H, Ph); other groups δ 1.87, 2.02 (2s, 6H, 2AcN), 2.06 (2), 2.15, 2.19 (2) (5s, 15H, 5AcO), and 6.24, 6.48 (2d, 2H, 2NH). Anal. Calcd for C42H60N2O20Si (941.0): C, 53.60; H, 6.43; N, 2.98. Found: C, 53.58; H, 6.45; N, 2.71.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4,7,8,9-tetra-Oacetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-2-acetamido-6-O-benzoyl-2-deoxy- β -D-galactopyranoside (14). Glycosylation of 9 (100 mg, 0.23 mmol) with 1 (250 mg, 0.48 mmol) in acetonitrile (3 mL) and tetrahydrofuran (3 mL) in the presence of DMTST (170 mg) and molecular sieves 3Å (670 mg), as described for 10, afforded 14 (98 mg, 46%) as an amorphous mass: [α]_D -30.5° (c 0.5, CHCl3); ¹H NMR (CDCl3) Neu5Ac unit δ 2.66 (dd, 1H, J_{3a,3e} = 12.5 Hz, J_{3e,4} = 4.4 Hz, H-3e), 3.71 (s, 3H, MeO), 4.80 (m, 1H, H-4), 5.26 (dd, 1H, J_{6,7} = 2.6 Hz, J_{7,8} = 9.5 Hz, H-7), and 5.71 (m, 1H, H-8); GalNAc unit δ 0.92 (m, 2H, Me3SiCH₂CH₂), 3.60 (m, 1H, Me3SiCH₂CH₂), and 7.40-8.05 (m, 5H, Ph); other groups δ 1.87, 2.03 (2s, 6H, 2AcN), 2.03, 2.14 (3) (4s, 12H, 4AcO), and 5.69, 5.94 (2d, 2H, NH).

Anal. Calcd for C40H58N2O19Si (899.0): C, 53.44; H, 6.50; N, 3.12. Found: C, 53.51; H, 6.73; N, 3.05.

A sample of 14 (70 mg, 0.078 mmol) was acetylated with acetic anhydride (1 mL)pyridine (2 mL), as described for 11, to give 15 (73 mg, quantitative) as an amorphous mass: ¹H NMR (CDCl₃) Neu5Ac unit δ 2.58 (dd, 1H, J_{3a,3e} = 12.8 Hz, J_{3e,4} = 4.4 Hz, H-3e), 3.74 (s, 3H, MeO), 4.34 (dd, 1H, J_{8,9'} = 2.2 Hz, J_{9,9'} = 11.7 Hz, H-9), 4.85 (m, 1H, J_{3a,4} = 12.5 Hz, H-4), 5.25 (dd, 1H, J_{6,7} = 2.4 Hz, J_{7,8} = 9.2 Hz, H-7), and 5.67 (m, 1H, H-8); GalNAc unit δ 1.00 (m, 2H, Me₃SiCH₂CH₂), 3.62 (m, 1H, Me₃SiCH₂CH₂), 4.48 (d, 1H, J_{1,2} = 7.7 Hz, H-1), 4.95 (broad s, 1H, H-4), and 7.34-8.04 (m, 5H, Ph); other groups δ 1.84, 2.00, (2s, 6H, 2AcN), 2.03, 2.12 (2), 2.14 (2) (5s, 15H, 5AcO).

Anal. Calcd for C42H60N2O20Si (941.0): C, 53.60; H, 6.43; N, 2.98. Found: C, 53.74; H, 6.33; N, 2.96.

ACKNOWLEDGMENT

This work was supported in part by a Grant-in-Aid (No. 03255207) for the Scientific Research on Priority Areas from the Ministry of Education, Science and Culture of Japan.

REFERENCES AND FOOTNOTES

1. Y. Ito and T. Ogawa, Tetrahedron, 46, 89 (1990).

- 2. K. Okamoto and T. Goto, Tetrahedron, 46, 5835 (1990).
- a) T. Murase, H. Ishida, M. Kiso, and A. Hasegawa, Carbohydr. Res., 184, c1 (1988); b) A. Hasegawa, H. Ohki, T. Nagahama, H. Ishida, and M. Kiso, Carbohydr. Res., 212, 277 (1991); c) A. Hasegawa, T. Nagahama, H. Ohki, K. Hotta, H. Ishida, and M. Kiso, J. Carbohydr. Chem., 10, 493 (1991).
- 4. a) P. Fügedi and P. J. Garegg, *Carbohydr. Res.*, **149**, c9 (1986); b) O. Kanie, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., **7**, 501 (1988).
- 5. a) P. Konradsson, U. E. Udodong, and B. Fraser-Reid, *Tetrahedron Lett.*, **31**, 4313 (1990); b) G. H. Veeneman, S. H. van Leevwen, and J. H. van Boom, *Tetrahedron Lett.*, **31**, 1331 (1990).
- 6. H. Prabhanjan, K. Aoyama, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, in press, and the references cited therein.
- K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmen, G. Noori, and K. Stenvall, J. Org. Chem., 53, 5629 (1988).
- 8. Y. Ogawa, M. Kitagawa, Y. Fujishima, M. Kiso, and A. Hasegawa, Agric. Biol. Chem., 53, 1025 (1989).
- 9. A. Hasegawa, M. Ogawa, H. Ishida, and M. Kiso, J. Carbohydr. Chem., 9, 393 (1990).
- 10. T. Murase, A. Kameyama, K. P. R, Kartha, H. Ishida, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8, 265 (1989).
- 11. Y. Ito and T. Ogawa, Tetrahedron Lett., 28, 6221 (1987).
- 12. A. Hasegawa, J. Nakamura, and M. Kiso, J. Carbohydr. Chem., 5, 11, 21 (1986).